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Total Synthesis and Antifungal Activity of 9-Methoxystrobilurin L as the Originally Proposed 1,4-Benzodioxan Structure

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Abstract—Total synthesis of both enantiomers of 9-methoxystrobilurin L as the originally proposed 1,4-benzodioxan structure was successfully achieved. The ^1H and ^{13}C NMR spectra of synthesized 9-methoxystrobilurin L were compared with those of a naturally-occurring sample. It was strongly indicated that naturally-occurring 9-methoxystrobilurin L has not the originally reported 1,4-benzodioxan structure but a 1,5-benzodioxepin structure, the same as previously reported 9-methoxystrobilurin K. Antifungal activities of the synthesized compounds toward several typical fungi were also examined, and they were less active than 9-methoxystrobilurin K. © 2001 Elsevier Science Ltd. All rights reserved.

Introduction

9-methoxystrobilurin L was isolated as a new analogue of β -methoxyacrylate antibiotics from mycelial culture of basidiomycete *Favolaschia pustulosa* by Xenova's group in 1996.¹ The compound exhibits strong antifungal activities toward several typical fungi by inhibiting a mitochondrial respiration pathway as other β -methoxyacrylates (MOAs) do.² In addition, the compound shows remarkable cytostatic activity toward human Burkitt's lymphoma derived cell lines (Jijoye) at very low concentration. The ^1H and ^{13}C NMR spectra of 9-methoxystrobilurin L were very similar to those of 9-methoxystrobilurin K (**2**) previously reported by Anke et al.;³ however, it was assigned as an original 1,4-benzodioxepin structure **1**. On the other hand, Blunt and Munro isolated a compound showing almost the same ^1H and ^{13}C NMR spectra as **1** or **2** from another culture in 1996, and claimed the 1,5-benzodioxepin structure **3** based on the assignment of the HMBC spectrum in CDCl_3 for these compounds.⁴ Recently, Anke and Steglich formally revised the structure of 9-methoxystrobilurin K to **3** by comparison of a synthesized 1,5-benzodioxepin substructure with the degradation product of natural strobilurin K.⁵ Further, a total synthesis

of 9-methoxystrobilurin K **3** was recently achieved and its 1,5-benzodioxepin structure was synthetically confirmed in our previous paper.⁶

According to these results, 9-methoxystrobilurin L also seemed to have the same 1,5-benzodioxepin structure; however, it should be confirmed synthetically for future study of structure–activity relationships. In addition, 1,4-benzodioxan structure **1** was also expected to show strong biological activities because strobilurin E (**4**) having a similar 1,4-benzodioxan structure indicated strong antifungal and cytostatic activities (Fig. 1).⁷ In this paper, we would like to report the asymmetric total synthesis of both enantiomers of 9-methoxystrobilurin L as the originally proposed 1,4-benzodioxan structure and its antifungal activities.

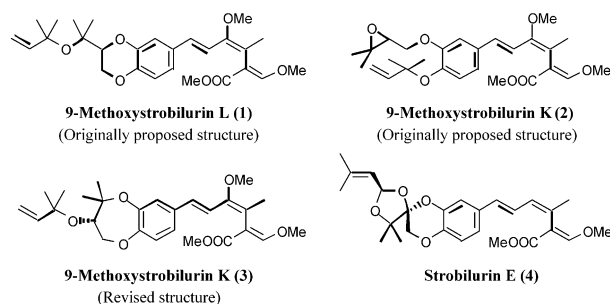


Figure 1. Cytostatic strobilurin analogues.

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Synthetic Plan

A retrosynthesis of 9-methoxystrobilurin L is shown in Scheme 1.

A construction of the relatively unstable conjugated triene moiety was to be performed at the last stage of total synthesis. Therefore, an enone-type intermediate **18** was decided as the first target molecule. Further, an optically active 1,4-benzodioxan **10** would be constructed by 6-*exo* selective intramolecular cyclization of epoxyphenol **9**. The epoxyphenol **9**, in turn, would be prepared from 2'-hydroxy-5'-bromoacetophenone **6** and chiral epichlorohydrin via Baeyer–Villiger oxidation.

Synthesis

The reaction of 2'-hydroxy-5'-bromoacetophenone **6** with optically active epichlorohydrin **5** was carried out by Augstein's α -selective coupling method⁸ and the desired ether **7** was obtained in 68% yield (Scheme 2).

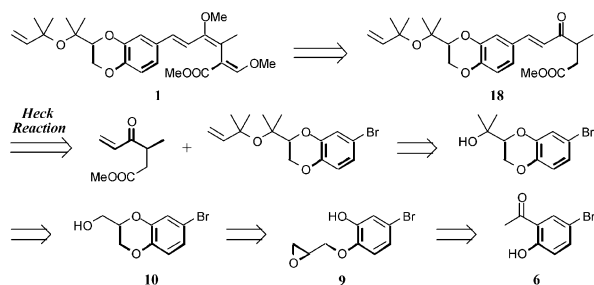
The Baeyer–Villiger oxidation of **7** proceeded smoothly to afford the corresponding epoxy ester **8**. Interestingly, the desired 6-*exo* cyclization subsequently proceeded on hydrolysis of the epoxy ester **8** for the preparation of cyclization precursor **9**. The thus-obtained primary alcohol **10** having a 1,4-benzodioxan-ring was oxidized to the carboxylic acid, which led to the corresponding methyl ester **11**. Treatment of the ester **11** with an excess

of methylmagnesium bromide gave a tertiary alcohol **12** in good yield. The optical purity of **12** was determined by HPLC analysis as 93% ee.⁹ A direct ether synthesis using the thus-obtained tertiary alcohol **12** with isoprene under acidic conditions failed due to the exclusive polymerization of the diene. Therefore, stepwise synthesis was next carried out. The α -alkoxy- α -methyl ester **13** was prepared by treatment of the alcohol **12** with sodium hydride and alkylation of the resulting alkoxide with methyl α -bromopropionate. Further, an enolate anion generated from **13** and sodium bis(trimethylsilyl)amide was methylated by the addition of methyl iodide. The prepared α,α -dimethyl- α -alkoxy ester **14** was reduced to the alcohol and subsequently oxidized to the corresponding aldehyde **15**. Terminal olefination of the aldehyde **15** utilizing Wittig reaction gave the aryl bromide **16**, and the desired tertiary–tertiary ether linkage was successfully constructed.

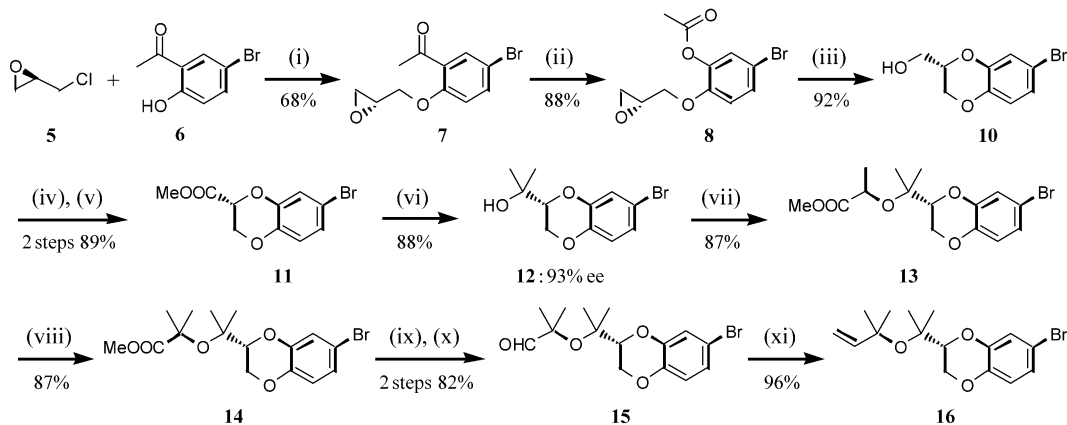
The Heck reaction of the aryl bromide **16** with a vinyl ketone **17**¹⁰ proceeded smoothly to give the desired enone-type intermediate **18** (Scheme 3). The ester **18** was hydrolyzed to the corresponding carboxylic acid, which led to the methylenolether **19** by treatment with potassium *tert*-butoxide and dimethyl sulfate. Further, the β -methoxyacrylate moiety was constructed by our previously reported method,¹⁰ and total synthesis of (*R*)-9-methoxystrobilurin L (**1R**) was successfully achieved. The enantiomer (**1S**) was also synthesized by the same procedure.

Spectral Comparison

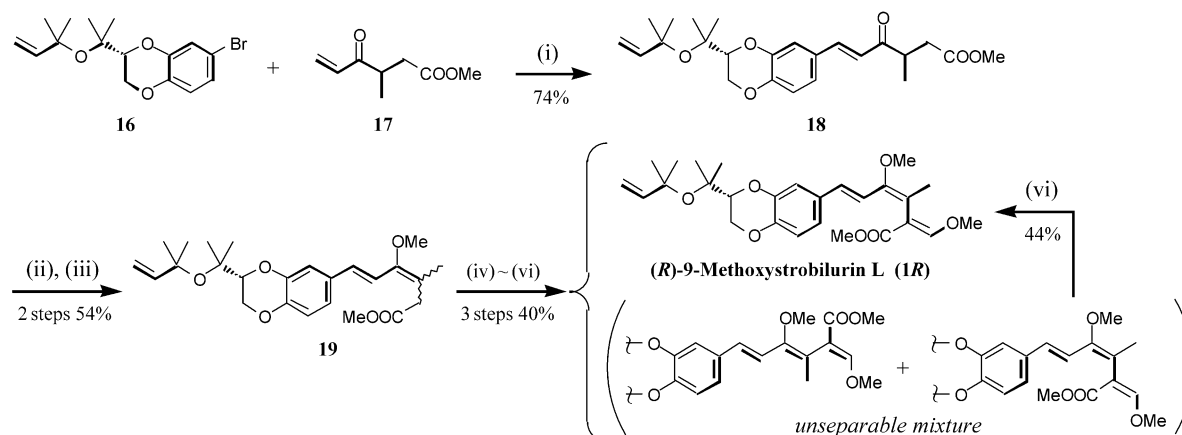
The ¹H and ¹³C NMR spectra of synthesized 9-methoxystrobilurin L (**1**)¹¹ were compared with those of a natural sample reported by Xenova's group^{1,13} (Typical ¹H NMR data are summarized in Table 1). The present study made it clear that 9-methoxystrobilurin L was not the 'originally reported' 1,4-benzodioxan structure because of the obvious differences in the chemical shifts on its ¹H and ¹³C NMR spectra in CD₃OD. On the other hand, a greater spectral similarity between natu-



Scheme 1. Retrosynthesis of 9-methoxystrobilurin L.

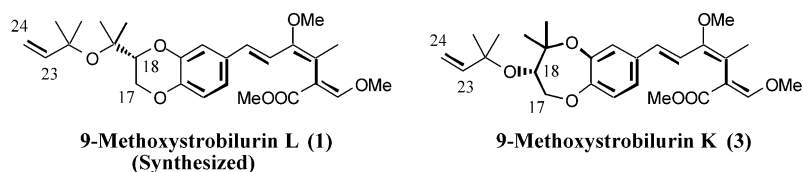


Scheme 2. Synthesis of 1,4-benzodioxan including tertiary–tertiary ether linkage (*R*-form). Reaction conditions: (i) KOH, EtOH–H₂O, reflux; (ii) mCPBA, CHCl₃, reflux; (iii) NaOH aq–THF, reflux; (iv) KMnO₄, KOH, H₂O, rt; (v) CN₂N₂, Et₂O, rt, 20 min; (vi) MeMgBr (excess), THF, rt, 1 h; (vii) NaH, methyl 2-bromopropionate, 0 °C; (viii) NaHMDS, MeI, THF, –78 °C; (ix) DIBAL, CH₂Cl₂, –78 °C; (x) (COCl)₂, DMSO, Et₃N, CH₂Cl₂, –60 °C; (xi) Ph₃P=CH₂, DMSO–THF, rt.



Scheme 3. Synthesis of (*R*)-9-methoxystrobilurin L. Reaction conditions: (i) 10 mol% Pd(OAc)₂, PPh₃, Et₃N, 100 °C; (ii) NaOH aq–MeOH, rt then HCl; (iii) KO^tBu, Me₂SO₄, DMF, –45 to –15 °C; (iv) NaH, HCOOMe, rt; (v) K₂CO₃, Me₂SO₄, HCOOMe, rt; (vi) hv (λ: 365 nm), acetone–benzene, rt.

Table 1. ¹H NMR spectra of 9-methoxystrobilurins L and K^{a,b}



Compound	9-Methoxystrobilurin L		9-Methoxystrobilurin K
	Natural	Synthesized	Synthesized
18	3.70 (m)	3.84 (dd, <i>J</i> = 2.0, 8.8 Hz)	3.69 (dd, <i>J</i> = 3.4, 7.6 Hz)
17a,17b	4.00 (dd, <i>J</i> = 7.5, 12.4 Hz)	3.97 (dd, <i>J</i> = 8.8, 11.2 Hz)	3.98 (dd, <i>J</i> = 7.6, 12.5 Hz)
	4.20 (dd, <i>J</i> = 3.1, 12.3 Hz)	4.55 (dd, <i>J</i> = 2.0, 11.2 Hz)	4.20 (dd, <i>J</i> = 3.4, 12.5 Hz)
23	5.90 (dd, <i>J</i> = 10.0, 17.4 Hz)	6.07 (dd, <i>J</i> = 10.7, 17.6 Hz)	5.93 (dd, <i>J</i> = 11.0, 17.7 Hz)
24a,24b	5.19 (dd, <i>J</i> = 0.7, 10.0 Hz)	5.00 (dd, <i>J</i> = 1.0, 10.7 Hz)	5.17 (dd, <i>J</i> = 0.9, 10.7 Hz)
	5.22 (dd, <i>J</i> = 0.7, 18.0 Hz)	5.13 (dd, <i>J</i> = 1.0, 17.6 Hz)	5.23 (dd, <i>J</i> = 0.9, 17.7 Hz)

^aCD₃OD was used as the solvent.

^bTetramethylsilane was used as the internal standard.

Table 2. Antifungal activities of synthesized (*R*)- and (*S*)-9-methoxystrobilurin L

Compound	Diameter of inhibition zone (mm) ^{a,b}					
	Conc. (μg/disk)	<i>Penicillium citrium</i>	<i>Aspergillus fumigatus</i>	<i>Fusarium solani</i>	<i>Candida albicans</i>	<i>Saccharomyces cerevisiae</i>
Nystatin (positive control)	10	10	12	—	16	17
(1R)	10	—	8	—	10	8
	1	—	±	—	±	±
	0.1	—	±	—	—	—
(1S)	10	—	8	—	8	8
	1	—	8	—	—	—
	0.1	—	±	—	—	—
(3)	10	—	10	25*	22	25
	1	—	8	19*	17	20
	0.1	—	±	—	—	±

^aThe diameter of each inhibition zone (mean values of two samples) was measured after 48 h incubation.

^b—, Not effective; ±, slightly effective; *, incomplete inhibition.

rally-occurring 9-methoxystrobilurin L and synthesized 9-methoxystrobilurin K¹² was clearly demonstrated.

Biological Assay

Antifungal activities of both synthesized enantiomers of 9-methoxystrobilurin L toward several typical fungi examined by the disk diffusion method using potato dextrose agar are shown in Table 2.

Interestingly, both synthesized enantiomers of 9-methoxystrobilurin L were unexpectedly less active than 9-methoxystrobilurin K and nystatin (positive control) toward all of the examined fungi.

Conclusion

The originally proposed 1,4-benzodioxan structure of 9-methoxystrobilurin L was successfully synthesized; however, its structural assignment was clearly ruled out by obvious differences in ¹H and ¹³C NMR spectra. In conclusion, it was strongly indicated that naturally-occurring 9-methoxystrobilurin L should be the same compound as 9-methoxystrobilurin K previously isolated by two other groups. In addition, both synthesized enantiomers of the 1,4-benzodioxan structure were less effective than 9-methoxystrobilurin K. This result is in contrast to the strong antifungal activity of strobilurin E (**4**) having a similar 1,4-benzodioxan structure and indicated that the geometry of the hindered ether-type side chain moiety on the 1,4-benzodioxan ring is very influential in the biological activities of these compounds. Further extensive studies for the synthesis and design of novel β-methoxyacrylate antibiotics having a more effective side chain moiety are now in progress.

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11. Physical data of synthesized 9-methoxystrobilurin L (**1**): ¹H NMR (δ, 500 MHz, CD₃OD) 1.31 (s, 3H), 1.36 (s, 6H), 1.44 (s, 3H), 1.84 (s, 3H), 3.65 (s, 3H), 3.71 (s, 3H), 3.81 (s, 3H), 3.78 (dd, 1H, *J*=2.0, 8.8 Hz), 3.94 (dd, 1H, *J*=8.8, 11.2 Hz), 4.48 (dd, 1H, *J*=2.0, 11.2 Hz), 5.00 (dd, 1H, *J*=1.0, 10.7 Hz), 5.13 (dd, 1H, *J*=1.0, 17.6 Hz), 6.12 (dd, 1H, *J*=10.7, 17.6 Hz), 6.34 (d, 1H, *J*=15.9 Hz), 6.54 (d, 1H, *J*=16.1 Hz), 6.74 (d, 1H, *J*=8.3 Hz), 6.81 (dd, 1H, *J*=2.0, 8.3 Hz), 6.84 (d, 1H, *J*=2.0 Hz), 7.48 (s, 1H); ¹³C NMR (δ, 125.6 MHz, CD₃OD) 16.5, 23.8, 26.9, 30.09, 30.11, 52.0, 59.8, 62.4, 66.4, 77.2, 78.1, 80.9, 111.5, 112.1, 115.5, 118.0, 120.7, 120.9, 128.5, 132.3, 144.8, 145.3, 148.7, 154.0, 161.4, 169.8; [α]_D²⁸=72.9 (**1R**), (*c*=0.402, CHCl₃), -71.5 (**1S**), (*c*=0.442, CHCl₃); HRMS calcd for C₂₇H₃₆O₇ (M⁺) 472.2461, found 472.2446 (**1R**), 472.2452 (**1S**).
12. Physical data of synthesized 9-methoxystrobilurin K (**3**): ¹H NMR (δ, 500 MHz, CD₃OD) 1.22 (s, 3H), 1.32 (s, 3H), 1.32 (s, 3H), 1.38 (s, 3H), 1.85 (s, 3H), 3.63 (s, 3H), 3.69 (dd, 1H, *J*=3.4, 7.6 Hz), 3.70 (s, 3H), 3.83 (s, 3H), 3.98 (dd, 1H, *J*=7.6, 12.5 Hz), 4.20 (dd, 1H, *J*=3.4, 12.5 Hz), 5.17 (dd, 1H, *J*=0.9, 10.7 Hz), 5.23 (dd, 1H, *J*=0.9, 17.7 Hz), 5.93 (dd, 1H, *J*=11.0, 17.7 Hz), 6.36 (d, 1H, *J*=15.9 Hz), 6.54 (d, 1H, *J*=15.9 Hz), 6.80 (d, 1H, *J*=8.2 Hz), 6.88 (d, 1H, *J*=2.1 Hz), 6.94 (dd, 1H, *J*=2.1, 8.2 Hz), 7.49 (s, 1H); ¹³C NMR (δ, 125.6 MHz, CD₃OD) 16.5, 22.6, 26.3, 26.7; [α]_D²⁴=-8.87 (*c*=0.860, CHCl₃); HRMS calcd for C₂₇H₃₆O₇ (M⁺) 472.2461, found 472.2456.
13. Physical data of natural 9-methoxystrobilurin L:¹ ¹H NMR (δ, 400 MHz, CD₃OD) 1.21 (s, 3H), 1.32 (s, 3H), 1.32 (s, 3H), 1.39 (s, 3H), 1.85 (s, 3H), 3.62 (s, 3H), 3.70 (s, 3H), 3.70 (m, 1H), 3.82 (s, 3H), 4.00 (dd, 1H, *J*=7.5, 12.4 Hz), 4.20 (dd, 1H, *J*=3.1, 12.3 Hz), 5.19 (dd, 1H, *J*=0.7, 10.0 Hz), 5.22 (dd, 1H, *J*=0.7, 18.0 Hz), 5.90 (dd, 1H, *J*=10.0, 17.4 Hz), 6.39 (d, 1H, *J*=15.0 Hz), 6.54 (d, 1H, *J*=15.0 Hz), 6.80 (d, 1H, *J*=8.4 Hz), 6.84 (d, 1H, *J*=2.1 Hz), 6.97 (dd, 1H, *J*=2.1, 8.2 Hz), 7.44 (s, 1H); ¹³C NMR (δ, 100 MHz, CD₃OD) 16.5, 22.6, 26.4, 26.8, 28.2, 52.2, 59.9, 62.6, 72.8, 77.4, 77.7, 82.9, 111.9, 114.9, 119.6, 121.3, 121.5, 122.9, 123.7, 128.3, 134.5, 145.3, 147.9, 152.2, 154.3, 161.6, 169.9.